

# Superparamagnetic cobalt ferrite nanoparticles as $T_2$ contrast agent in MRI: in vitro study

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**Abstract:** Superparamagnetic cobalt ferrite nanoparticles ( $\text{CoFe}_2\text{O}_4$ ) possess favourite advantages for theranostic applications. Most of previous studies reported that  $\text{CoFe}_2\text{O}_4$  magnetic nanoparticles (MNPs) are suitable candidates for induction of hyperthermia and transfection agents for drug delivery. The present study synthesized and investigated the potential use of  $\text{CoFe}_2\text{O}_4$  as a contrast agent in magnetic resonance imaging (MRI) by using a conventional MRI system. The  $\text{CoFe}_2\text{O}_4$  were synthesized using co-precipitation method and characterized by TEM, XRD, FTIR, EDX and VSM techniques. Relaxivities  $r_1$  and  $r_2$  of  $\text{CoFe}_2\text{O}_4$  were then calculated using a 1.5 Tesla clinical magnetic field. The cytotoxicity of  $\text{CoFe}_2\text{O}_4$  was evaluated by the MTT assay. Finally, the optimal concentrations of MNPs for MRI uses were calculated through the analysis of  $T_2$  weighted imaging cell phantoms. The superparamagnetic  $\text{CoFe}_2\text{O}_4$  NPs with an average stable size of 10.45 nm were synthesized. Relaxivity  $r_{1,2}$  calculations resulted in suitable  $r_2$  and  $r_2/r_1$  with values of 58.6 and 51 that confirmed the size dependency on relaxivity values. The optimal concentration of MNPs for MR image acquisition was calculated as 0.154 mM. Conclusion:  $\text{CoFe}_2\text{O}_4$  synthesized in this study could be considered as a suitable  $T_2$  weighted contrast agent because of its high  $r_2/r_1$  value.

## 1 Introduction

Magnetic nanoparticles (MNPs) have been widely employed for various applications including magnetic resonance imaging (MRI), cancer hyperthermia, drug delivery, tissue imaging [1, 2]. Ferrite-based MNPs have been widely-explored as magnetic nanomaterials because of their excellent magnetic properties and multifunctional agents [3, 4]. Because of superparamagnetic properties of some ferrite-based nanoparticles, they have been largely employed to enhance the proton relaxivity for improved contrast and sensitivity of MR image acquisition [5]. In addition, ferrite-based MNPs could enhance the efficiency of hyperthermia owing to their high anisotropy, making them suitable candidates for theranostic applications. [6]. Among various ferrite-based MNPs, cobalt ferrite ( $\text{CoFe}_2\text{O}_4$ ) nanoparticles have been recognised as a favourite contrast agent. They have hard magnetic material properties such as high saturation magnetisation, strong anisotropy and mechanical hardness [7]. Size and nanostructure of MNPs affect their magnetic properties that could be highly modulated by the preparation methods [8]. Water-soluble ferrite MNPs could be synthesised by a co-precipitation method for the biological applications [9]. Ferrite-based MNPs are used as contrast agents for  $T_2$ -weighted MR images. Based on theoretical models and experimental reports,  $r_2$  relaxivity depends on the particles size and aggregation, the square of the saturation magnetisation and the applied magnetic field strength [10]. For small MNPs, which satisfy the motional averaging regime (MAR),  $\Delta\omega\tau_D < 1$ , the outer sphere theory ( $R = (16/45)f(\Delta\omega)\tau_D\alpha d^2Hyd$ ) could be applied. This theory is not applicable for larger sizes of MNPs. The static dephasing regime is dominant for too large sizes. In this regard,  $r_2$  enhances as size increases [2, 11]. According to Ta *et al.* [12]  $r_2$  relaxivity increased as magnetic field strength enhanced from 1.5 to 9.4 T and then decreased.  $r_2/r_1$  is an indicator of MRI efficiency that

increases with the increment of field strength [13]. Some studies evaluated  $\text{CoFe}_2\text{O}_4$  MNPs as MR contrast agent in different sizes and various magnetic fields, confirming the above-mentioned content [2, 14–17]. The studies indicated that high  $r_2/r_1$  was obtained in large sizes and high magnetic fields. For clinical applications of superparamagnetic  $\text{CoFe}_2\text{O}_4$ , it is necessary to obtain high  $r_2$  and  $r_2/r_1$  in conventional MRI systems. A contrast agent with optimal efficiency within the range of clinical magnetic field might find clinical applications. In this study, the superparamagnetic  $\text{CoFe}_2\text{O}_4$  MNPs were synthesised with high size stability, and characterised by high  $r_2/r_1$  in clinical magnetic field strength. In addition, the cytotoxicity of this multifunctional agent was evaluated.

## 2 Material and methods

### 2.1 Materials

Co (II) and Fe (III) were purchased from Aldrich, Scharlau and Alfa Aesar. NaOH was obtained from Merck. MTT (3-(4, 5-Dimethylthiazol-2-yl)-2, 5-diphenyltetrazolium Bromide) was purchased from Sigma Aldrich. Agarose gel and deionised water (DI water) were used during the tests.

### 2.2 Preparation of $\text{CoFe}_2\text{O}_4$ MNPs

$\text{CoFe}_2\text{O}_4$  MNPs were synthesised by co-precipitation method in an alkaline aqueous environment. The reaction mixture was prepared from iron sulphate ( $\text{Fe}_2(\text{SO}_4)_3$  salts) and cobalt chloride ( $\text{CoCl}_2$  salt) with 0.1 M concentration of metal salts. All components of the reaction mixture were deoxygenated with nitrogen gas before mixing. In the next step, 5.0 M NaOH solution was added with vigorously stirring of mixing reaction until reaching a pH of 12.4. The obtained solution was then replaced while stirring at 80°C for

3 h under continuous nitrogen gas bubbling. Finally, the obtained sedimentary solution was centrifuged at 8500 rpm for 3 min, and was carefully rinsed 3 times using 10 ml of DI water. The sediment was then rinsed with ethanol. Afterwards,  $\text{CoFe}_2\text{O}_4$  MNPs were dried at 50°C in dry heat [18].

## 2.3 Characterisation

**2.3.1 Transmission electron microscopy (TEM):** TEM was carried out to evaluate the morphology and size distribution of the synthesised particles. A 200 keV field emission Tecnica F 20 (FE) TEM was used to get a high-resolution TEM (HRTEM) and selected area (electron) diffraction (SAED) pattern. The particle size histogram was determined by measuring the diameter of ~508 NPs and fitted by a Gaussian distribution.

**2.3.2 Scanning electron microscopy (SEM):** SEM was carried out to evaluate the morphology and surface structure of the MNPs. A 20 kV High Voltage MIRA3 TESCAN in 1.38  $\mu\text{m}$  of the field of view was used for SEM.

**2.3.3 Phase structure:** The X-ray diffraction (XRD) indicated  $\text{CoFe}_2\text{O}_4$  MNPs with crystal lattice structures. The data was collected at the room temperature on an X-ray diffractometer (GMR EXPLORER, ITALY). The Pure  $\text{CoFe}_2\text{O}_4$  MNPs was obtained at a calcination temperature of 550°C [19]. The XRD system was run at 40 kV and 30 mA in a  $2\theta$  range of 20°–80°. In the present study, dimensions of  $\text{CoFe}_2\text{O}_4$  MNPs crystal ( $D$ ) were estimated using the XRD information through the Sherrer's equation.

**2.3.4 Infrared spectra:** Fourier-transform IR (FTIR) spectroscopy was used to identify functional groups and chemical structural changes in materials. For preparation of sample: the powder sample and KBr salt were ground to reduce the particles size. Then, a small amount of powder sample (about 0.1–2% of the KBr amount) was mixed with the KBr powder. Subsequently, the mixture was ground for 3–5 min. Uniformly fine-grained powders were prepared using milling the mixture using a mechanical vibrator or a mill. A thin and transparent pellet was obtained under pressure. Then, the pellet was put onto the sample holder in the FTIR system. The infrared spectrum was recorded by FTIR spectrometer (AVATAR 370 FT-IR Thermo Nicolet Spectrum) that operated at room temperature. It was performed by 64 scans and the samples were analysed in transmittance mode [20]. The Spectral resolution of the system was set at 4  $\text{cm}^{-1}$  [21].

**2.3.5 Energy dispersive X-Ray spectroscopy (EDS or EDX) spectrum:** EDX spectrum indicated the presence of Fe, Co and O elements for the elemental analysis or chemical characterisation of samples.

**2.3.6 Magnetometry:** A vibrating sample magnetometer (vibrating sample magnetometer) VSM (manufactured by Danesh Pajoush Magnetis Company of Kashan, VSMF model, Iran) was used to measure the magnetic field-dependent magnetisation loop from –15,000 to 15,000 Oe at room temperature.

## 2.4 Relaxometry

Contrast agents in MRI could change relaxation times in tissues of interest. They can reduce  $T_1$  and  $T_2$  relaxation times that can be introduced as positive or negative contrast agents in MR images, respectively.  $\text{CoFe}_2\text{O}_4$  MNPs are often used to enhance  $T_2$  contrast are referred to as  $T_2$  weighted images.  $T_2$  and  $r_2$  of water protons through the synthesised  $\text{CoFe}_2\text{O}_4$  MNPs were calculated at 1.5 T MRI scanner (Avanto/Siemens, Kamyab Hospital). An in vitro phantom containing  $\text{CoFe}_2\text{O}_4$  MNPs with various concentrations of 0.03, 0.04, 0.8, 0.12, 0.21, 0.25, 0.31, 0.36 and 0.42 mM was used to measure  $r_1$  and  $r_2$  values. All curve fitting routines, which were used to determine relaxation rate maps, were performed by Excel

and R Software.  $T_1$ -weighted image was acquired at TE: 8.7 ms; TR1/TR2/TR3/TR4/TR5/TR6: 100/300/600/900/1200/2000 ms; flip angle: 20°; matrix:  $256 \times 192$ ; the field of view: 260 mm; 100%; averages: 1, echo train length: 1; slice thickness: 5 mm.  $T_2$ -weighted images were obtained by a  $T_2$  spin-echo multisection pulse sequence with fixed repetition time (TR) of 2000 ms; TE1/TE2/TE3/TE4/TE5/TE6/TE7/TE8/TE9/TE10/TE11/TE12/TE13/TE14/TE15/TE16: 13.8/27.6/41.4/55.2/69/82.8/96.6/110.4/124.2/138/151/165.6/179.4/193.2/207/220.8, flip angle: 20°; matrix:  $256 \times 192$ ; field of view: 260 mm; 100%; averages: 1, echo train length: 1.

## 2.5 In-vitro MR imaging of cell phantoms

In vitro experiments were performed using KYSE 30 (RRID: CVCL1351), an oesophagus cancer cell line (CCLE) from Homo sapiens (Human), extracted from a 64-year-old man. KYSE 30 cells were seeded at a density of  $2 \times 10^6$  at T12.5 culture flasks. After 24 h, different concentrations of  $\text{CoFe}_2\text{O}_4$  composite (0.04, 0.8, 0.12, 0.21, 0.25, 0.31 mM) were added to the culture flasks. Culture flasks were then rinsed by PBS. The cells were then detached and centrifuged at a microtube (2 cc). Few drops of agarose gel were added to every microtube to fix cells followed by sonication to remove air bubbles. MR imaging of cell phantoms was performed for two times using a 1.5 T MRI system (Avanto/SIEMENS, KAMYAB HOSPITAL). A  $T_2$ -Tse-cor gradient-echo sequence was acquired using the following sequence parameters: TR: 4000 ms, TE: 81 ms; flip angle: 20°; matrix:  $256 \times 192$  interpolated; the field of view: 260 mm; averages: 1, echo train length: 1; slice thickness: 8 mm; 4 slices. The signal intensity (SI) was obtained from different concentrations of MNPs using Radiant Software (4.6.8 evaluation version) in cell phantoms. Percentage of  $\Delta\text{SI}$  ( $(\text{SI}/\text{SI}_{\text{Control}}) \times 100$ ) was calculated in which  $\text{SI}_{\text{Control}}$  refers to cell phantom without MNPs.

## 2.6 Cytotoxicity of $\text{CoFe}_2\text{O}_4$ MNPs

KYSE 30 was seeded at a density of  $10^4$  cells/well in a 96 well plate and incubated at 37°C for a doubling time of KYSE30 cell line for sufficient growth.  $\text{CoFe}_2\text{O}_4$  MNPs in concentrations of 0.05, 0.1, 0.2, 0.3, 0.4, 0.5, 0.75 and 1 mM were separately added to microwells. Cells were then incubated for 24 h. Finally, MTT (3-(4, 5-Dimethylthiazol-2-yl)-2, 5-diphenyltetrazolium Bromide) test was performed to determine the cell death percentage.

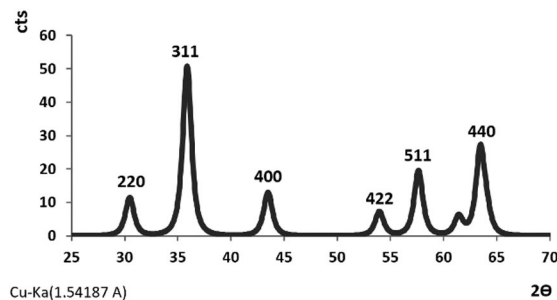
## 3 Results

### 3.1 XRD pattern

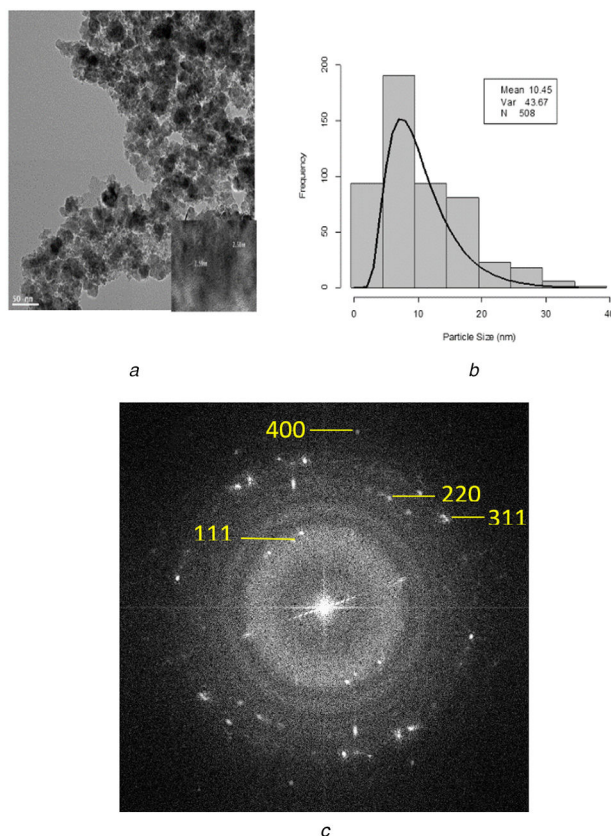
Fig. 1 shows the obtained XRD result of  $\text{CoFe}_2\text{O}_4$  MNPs. The XRD profile from  $\text{CoFe}_2\text{O}_4$  MNPs, prepared by co-precipitation method, revealed the maximum XRD peak occurred at  $2\theta$  value of 35.87° that represented a typical  $\text{CoFe}_2\text{O}_4$  with an interlayer spacing value of 3.83242 Å. The structural analysis of XRD pattern indicated that  $\text{CoFe}_2\text{O}_4$  MNPs had an inverse cubic spinel-type. Mean size of crystals ( $D$ ) was estimated by the Sherrer's equation:  $D = K\lambda/\beta \cos\theta$  where  $K$  is the Scherrer constant (0.94),  $\lambda$  is the wavelength,  $\beta$  is the FWHM (in radians), and  $\theta$  is the peak angular position. Consequently, the size of the  $\text{CoFe}_2\text{O}_4$  crystal was calculated by the most intensive peak (311) with a value of 11.67 nm [1, 2, 22–24]. The peaks of (111), (220), (311), (400), (511), and (440) were the main peaks of the typical inverse cubic  $\text{CoFe}_2\text{O}_4$  MNPs XRD spectrum [25–28]. The peak (311) was used to obtain the lattice constant ( $a$ ) of  $\text{CoFe}_2\text{O}_4$  MNPs according to the following equation [29]:

$$a = d_{hkl} \sqrt{h^2 + k^2 + l^2}$$

where  $d_{hkl}$  is an interplanar distance;  $h$ ,  $k$  and  $l$  refer to Miller indices and the lattice constant. The constant ( $a$ ) of  $\text{CoFe}_2\text{O}_4$



**Fig. 1** XRD pattern of sample  $\text{CoFe}_2\text{O}_4$  synthesised by a co-precipitation method. The XRD system acted at 40 kV and 30 mA in a  $2\theta$  range of 20–80 [26, 30]



**Fig. 2** HRTEM images of  $\text{CoFe}_2\text{O}_4$  MNPs

(a) HRTEM of  $\text{CoFe}_2\text{O}_4$  MNPs, (b) Size distribution histogram of  $\text{CoFe}_2\text{O}_4$  MNPs with Gaussian distribution, (c) SAED pattern of  $\text{CoFe}_2\text{O}_4$  MNPs [31, 32]

crystal was computed according to the peak (31 1) with a value of 0.83 nm.

### 3.2 High-resolution TEM

Fig. 2a shows the HRTEM image of  $\text{CoFe}_2\text{O}_4$  MNPs. It is clear that  $\text{CoFe}_2\text{O}_4$  MNPs synthesised were aggregated and had non-uniform shapes. Fig. 2b shows the nanoparticle size distribution in the histogram. Size distribution was determined by measuring the mean diameter of about 508 particles on HRTEM image. The average size was 10.45 nm that was fitted by a Gaussian distribution. The SAED pattern (Fig. 2c) shows at least four well-defined diffraction rings. The rings were indexed with estimating their d-spacing as (111), (311), (220) and (400) reflections of the cubic  $\text{CoFe}_2\text{O}_4$  MNPs that are in agreement with the XRD results.

### 3.3 SEM

Fig. 3 shows the SEM image of  $\text{CoFe}_2\text{O}_4$  MNPs that used to confirm the morphology of the synthesised MNPs. The obtained results show that  $\text{CoFe}_2\text{O}_4$  MNPs had non-uniform shape. The average size was estimated as 25.6 nm using SEM analysis. Size estimation was performed by ImageJ software.

### 3.4 VSM

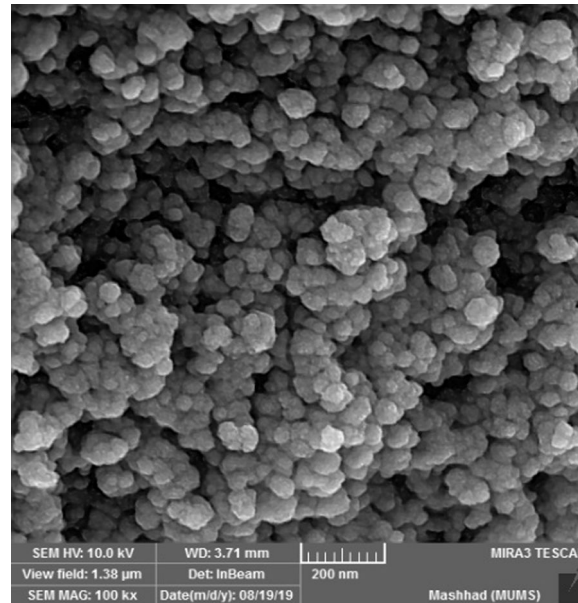
Magnetic properties of the synthesised  $\text{CoFe}_2\text{O}_4$  MNPs were evaluated using a SQUID system. Magnetic hysteresis curves of MNPs were obtained at the magnetic field within the range of  $-15,000$ – $15,000$  Oe at room temperature. The magnetisation of  $\text{CoFe}_2\text{O}_4$  MNP samples was not saturated by the SQUID (X Fig. 4). Therefore, the saturation magnetisation was obtained through extrapolation with a value of 7.5 emu/g [33].

### 3.5 EDX spectrum

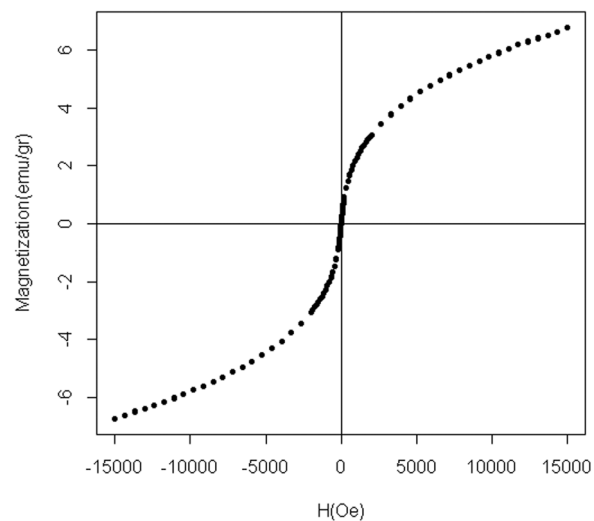
EDX spectrum of  $\text{CoFe}_2\text{O}_4$  MNPs shows the presence of elements of Fe, Co and O (Fig. 5). The peaks in the EDX pattern were perfectly assigned to the elements present in  $\text{CoFe}_2\text{O}_4$  nanoparticles.

### 3.6 FTIR spectra

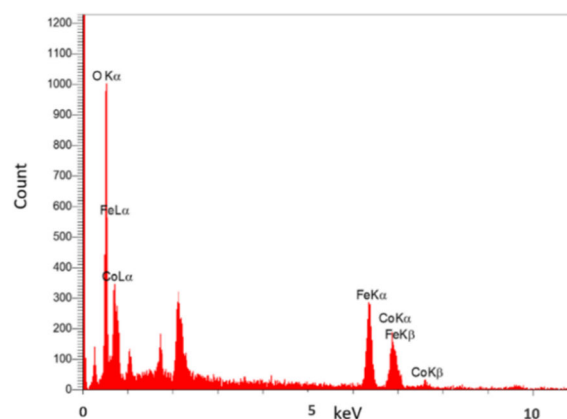
Fig. 6 shows the FTIR of  $\text{CoFe}_2\text{O}_4$  MNPs. Two main bands at  $590.31$  and  $416.35$   $\text{cm}^{-1}$  are assigned to M–O bond in octahedral and tetrahedral sites.  $416.3$   $\text{cm}^{-1}$  is related to Co–O band and  $590.3$   $\text{cm}^{-1}$  is associated with Fe–O band that confirmed the formation of  $\text{CoFe}_2\text{O}_4$  in the sample. The FT-IR spectra showed



**Fig. 3** SEM images of  $\text{CoFe}_2\text{O}_4$  MNPs. Size estimation was performed by imageJ software



**Fig. 4** Magnetic hysteresis curves for  $\text{CoFe}_2\text{O}_4$  MNPs. It was obtained at the magnetic field in the range of  $-15,000$  to  $15,000$  Oe at the room temperature



**Fig. 5** Energy dispersive X-ray spectroscopy (EDS or EDX) profile obtained from  $\text{CoFe}_2\text{O}_4$  MNPs

board bands at  $3380\text{ cm}^{-1}$  which are related to the OH group on the surface of nanoparticles. The bands at  $3414.3$  and  $1349.9\text{ cm}^{-1}$  are assigned due to the stretching of H–O–H bindings [34].

### 3.7 Relaxivity $r_2$

The MR capability of  $\text{CoFe}_2\text{O}_4$  MNPs was tested using a 1.5 T MRI system.  $T_1$  (longitudinal relaxation) and  $T_2$  (transverse relaxation) are two independent relaxation processes to generate an MR image. In the presence of MNPs, the relaxation rate ( $R = 1/T_{1,2}$ ) increases linearly with the MNPs concentration according to the following equation:

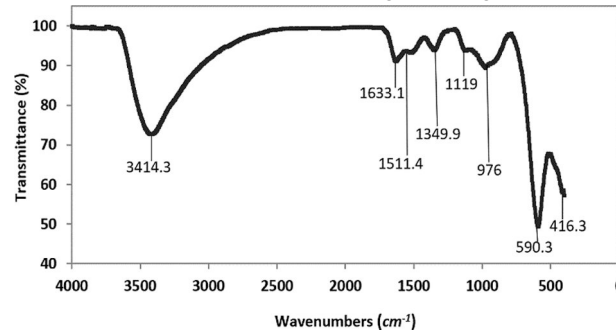
$$\frac{1}{T_{1,2}} = \frac{1}{T_0} + r_{1,2}C$$

where  $1/T_0$  is the relaxation rate of pure water and  $C$  is the concentration of MNPs [35].  $T_2$  spin-echo multisection pulse sequence with fix TR of 2000 ms and different TEs ranging between 13.8 and 220 ms were acquired for  $T_2$  measurement. Suspensions of  $\text{CoFe}_2\text{O}_4$  MNPs at various concentrations (0.03–0.42 mM) were prepared in the DI water in 2 ml microtubes and DI water acted as controls in experiments.  $T_2$  relaxation rates ( $1/T_2$ ) were obtained by analysing the TE-dependent SI curve for various concentrations of  $\text{CoFe}_2\text{O}_4$  MNPs (Fig. 7).  $T_2$  weighted image of

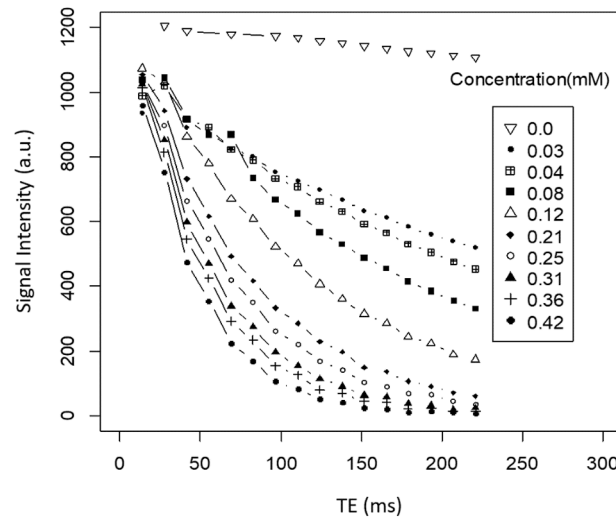
this nanostructure showed noticeable darkening by changing concentrations of MNPs in DI water. Thus, the SI of samples decreased at higher  $\text{CoFe}_2\text{O}_4$  MNPs concentrations (Fig. 8a). The  $r_2$  relativity was calculated as  $58.6 \text{ mM}^{-1} \text{ s}^{-1}$  according to the linear plot slope of the  $\text{CoFe}_2\text{O}_4$  MNPs concentration depending on the inverse  $T_2$  with  $R = 0.99$  (Fig. 8b).

### 3.8 Relaxivity $r_1$

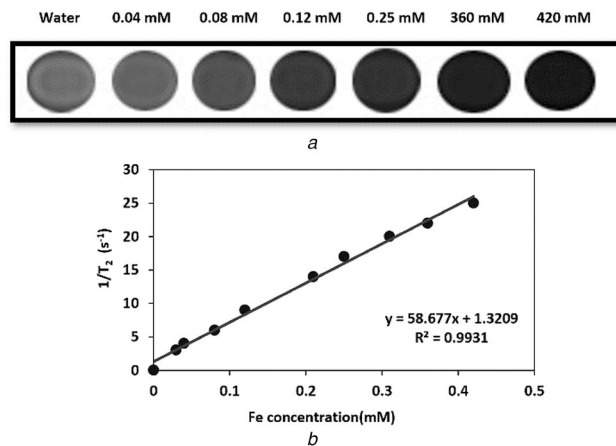
Using the same token,  $r_1$  was calculated by spin-echo with fix TE of 8.7 ms value and changing TR (from 100 to 2000 ms).  $T_1$  weighted image of this MNPs showed low darkening by changing



**Fig. 6** FTIR spectroscopy of  $\text{CoFe}_2\text{O}_4$  MNPs

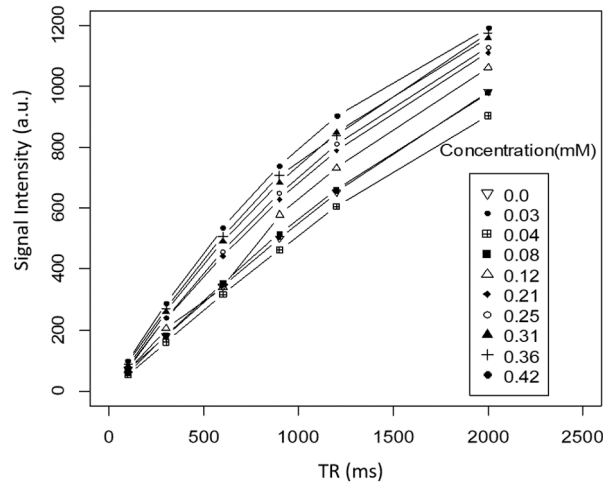


**Fig. 7** SI as a function of time of echo (TE) in various concentration of  $\text{CoFe}_2\text{O}_4$  MNPs. Spin-echo multisection pulse sequence with fix time of repetition (TR) of 2000 ms and different TEs ranging between 13.8 and 220 ms for  $T_2$  measurement at various concentrations (0.03–0.42 mM) that were prepared in the DI water and DI water played roles as a control sample

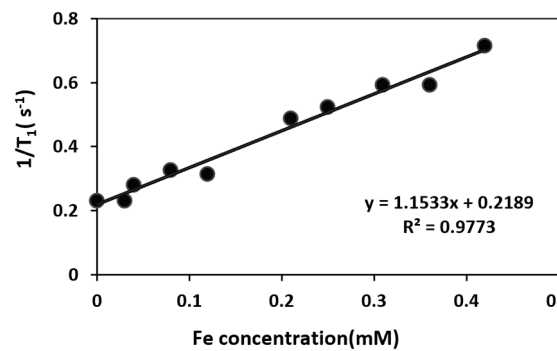


**Fig. 8** MR Image and calculated  $T_2$  relaxation rates and relaxivity

(a)  $T_2$ -weighted MR image of  $\text{CoFe}_2\text{O}_4$  MNPs in water medium obtained by a conventional spin-echo pulse sequence on a 1.5 T MRI system, (b)  $T_2$  relaxation rates ( $1/T_2$ ) depending on the concentration, calculated  $T_2$  relaxivity ( $r_2$ ) at various Fe concentrations of (0.03 to 0.42 mM)



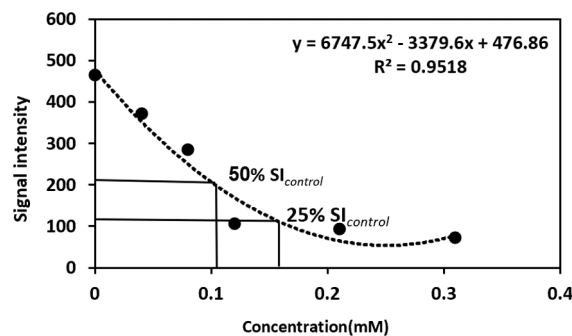
**Fig. 9** Spin echo sequence with fixed time of echo ( $TE$ ) of 8.7 value and changing time of repetition ( $TR$ ) (from 100 to 2000 ms) for  $T_1$  measurement at various concentrations (0.03–0.42 mM) of  $\text{CoFe}_2\text{O}_4$  MNPs that were prepared in the DI water



**Fig. 10** Calculated  $T_1$  relaxation rates and relativity at 1.5 T with varying Fe concentration of  $\text{CoFe}_2\text{O}_4$  MNPs (0.03 to 0.42 mM)

**Table 1** Changes in SI and percentage of  $\Delta SI$  ( $(SI/SI_{\text{control}}) \times 100$ ) which  $SI_{\text{control}}$  was related to cell phantom without MNPs) with concentration in cell phantoms image

| Concentration, mM | Signal intensity | $\Delta SI$ , % |
|-------------------|------------------|-----------------|
| 0                 | 464              | —               |
| 0.04              | 372              | 20              |
| 0.8               | 284              | 38              |
| 0.12              | 105              | 77              |
| 0.21              | 93.5             | 79              |
| 0.31              | 71.3             | 84              |



**Fig. 11** SI depended on concentration  $T_2$ - weighted MR image. 50 and 75%  $SI_{\text{control}}$  accrue at concentrations of 0.87 and 0.154 mM, respectively

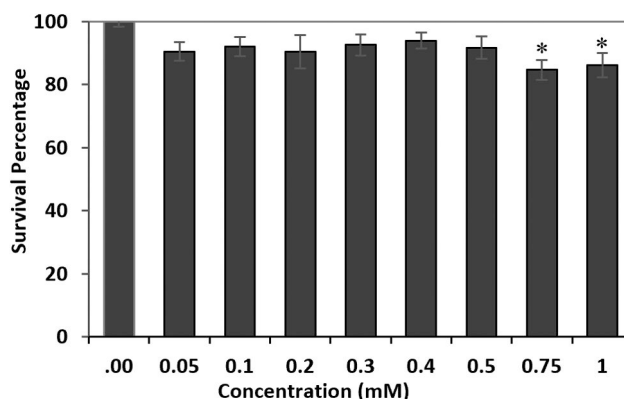
concentrations of this MNPs (Fig. 9). According to Fig. 10, the longitudinal relaxation rate,  $r_1$ , was  $1.15 \text{ mM}^{-1}\text{s}^{-1}$  for  $\text{CoFe}_2\text{O}_4$  MNPs in DI water.

$r_2/r_1$  ratio: The  $r_2/r_1$  ratio is an interesting sensitive parameter that is used to identify the category of the contrast agents ( $T_1$  or  $T_2$  contrast agent). The  $r_2/r_1$  ratio was calculated as 51 in the present study.

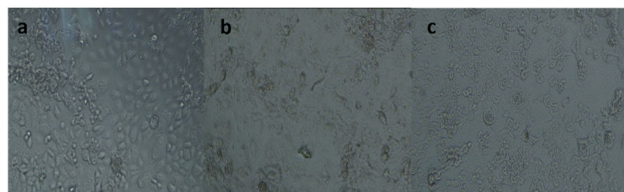
### 3.9 In vitro MR imaging of cell phantom

To optimise the clinical application of  $\text{CoFe}_2\text{O}_4$  MNPs, SI and  $\Delta SI$  ( $(SI/SI_{\text{control}}) \times 100$ ) were obtained from cell phantom  $T_2$  weighted image (X Table 1). The results indicated that the optimal concentration of  $\text{CoFe}_2\text{O}_4$  MNPs was 0.154 mM to obtain 75% of maximum decay (Fig. 11).





**Fig. 12** *In vitro* cytotoxicity of  $\text{CoFe}_2\text{O}_4$  MNPs tested on KYSE 30 cell line for 24 h by MTT assay. The \* sign indicates the significance of the statistic test



**Fig. 13** Morphology image of KYSE 30 cell

(a) Without treatment (control), (b) Treated by 0.75 mM of  $\text{CoFe}_2\text{O}_4$  MNPs, (c) Treated by 0.1 mM of  $\text{CoFe}_2\text{O}_4$  MNPs

**Cytotoxicity of  $\text{CoFe}_2\text{O}_4$  MNPs:** The cytotoxicity of  $\text{CoFe}_2\text{O}_4$  MNPs was evaluated by analysing the cell survival MTT assay using KYSE30 cells.  $\text{CoFe}_2\text{O}_4$  MNPs were incubated at various concentrations within the range of 0.01–1 mM for 24 h. The test showed a survival rate of more than 80% for the maximum concentration of 1 mM (Fig. 12). Statistical analysis was performed by One-Way ANOVA test to compare control cells with the treated cells. Significant  $p$ -value was obtained by 0.009 and 0.028 for 0.75 and 1 mM of the MNPs concentrations, respectively. Morphology images of the treated KYSE30 cells with various concentrations of  $\text{CoFe}_2\text{O}_4$  MNPs are shown in Fig. 13.

## 4 Discussion

This study aimed to synthesise and evaluate the performance of using superparamagnetic  $\text{CoFe}_2\text{O}_4$  MNPs as a suitable  $T_2$  contrast in clinical magnetic field strengths. Small size  $\text{CoFe}_2\text{O}_4$  MNPs were synthesised through co-precipitation method that could be the result of using a good multifunctional agent. Relaxometry of  $\text{CoFe}_2\text{O}_4$  MNPs, the optimal concentration of MNPs for MR imaging, and cytotoxicity effects of the MNPs were investigated.

HRTEM image of the synthesised  $\text{CoFe}_2\text{O}_4$  MNPs shows a non-uniform and heterogeneous morphology. Also, there was an aggregation. Previous studies reported that smaller particles have a higher aggregation tendency due to lower energy barriers [36, 37]. The size distribution obtained from the HRTEM image revealed an average size of 10.45 nm for the synthesised MNPs. This size range was chosen for two reasons. Firstly, the superparamagnetic nanoparticles were characterised by size of <20 nm [38]. Secondly, MNPs smaller than 7 nm could not positively affect the  $r_2$  relaxivity. However, slightly higher sizes could be optimal for the enhancement of the  $r_2$  relaxivity [15, 17].

SAED pattern revealed that the synthesised sample is in polycrystalline nature (Fig. 2c). It shows dotted rings pattern corresponding to spinel cubic  $\text{CoFe}_2\text{O}_4$  MNPs [31, 39, 40].

According to the SEM image, the synthesised MNPs showed non-uniform shape. Agglomeration of the MNPs could be the reason for higher size in SEM image that could be due to Van der Waals forces between the particles [41].

Through the analysis of XRD pattern, the crystal size of the synthesised  $\text{CoFe}_2\text{O}_4$  MNPs (D) was estimated by the Scherrer's equation with a small value of 11.67 nm. Subsequently, the lattice constant was estimated at 0.834 nm.

This finding is in agreement with the studies of Kalamet *et al.* [42] and Houshiar *et al.* [1] indicating that the constant (a) of nanocrystalline depends on the size and synthesis method.

According to the hysteresis curve analysis of the  $\text{CoFe}_2\text{O}_4$  MNPs, the lack of  $H_c$  and  $M_r$  refer to the superparamagnetic properties of the sample [33]. The small size effect and increased surface area of nanoparticles leading to superparamagnetic properties and the hysteresis curve without any loop [43]. The saturation magnetisation ( $M_S$ ) of small-sized MNPs can be described by a magnetic-dead layer model.

It can be explained by the demagnetisation of surface spin due to the surface-to-volume ratio effect. According to this model, the reduced MNP size increased the surface-to-volume ratio, and thus the increased dead layer component decreased  $M_S$  [15].

After the characteristics of MNPs, MR images of  $\text{CoFe}_2\text{O}_4$  MNPs were performed in an aqueous environment to evaluate the contrast between different concentrations of MNPs.  $T_2$  weighted imaging of MNPs show a noticeable contrast by changing concentrations of MNPs. Therefore,  $\text{CoFe}_2\text{O}_4$  MNPs could be considered as a negative contrast agent. Dephasing of the magnetic moment of protons could be caused by inhomogeneity in the magnetic field of environmental molecules in the presence of MNPs [44].

The  $r_2$  value of  $\text{CoFe}_2\text{O}_4$  MNPs was estimated at 58.6 by a magnetic relaxometry of  $\text{CoFe}_2\text{O}_4$  MNPs suspension at a 1.5 T conventional MRI system. The  $r_2$  value highly depends on size,  $M_S$  and the magnetic field strength [6]. The obtained  $r_2$  was consistent with reports by Kanget *et al.* indicating that  $T_2$  relativity depends on the particle size, mass magnetisation and the concentration of MNPs [2, 45, 46]. The result was consistent with the study performed by Joos *et al.* [11] who proved that smaller MNPs,  $r_2$  and  $r_2^*$  decreased in a higher range of particle size because of the high surface spin anisotropy [47]. It was also consistent with the MAR theory. The size-dependent effect was strongly observable at low frequencies [15]. Consequently, the value of  $r_2$  is related to small synthesised  $\text{CoFe}_2\text{O}_4$  MNPs, low frequency and  $M_S$  that depended on the magnetic-dead layer. Despite these results, Venkatesha *et al.* [48] showed that with decreasing the size of  $\text{CoFe}_2\text{O}_4$ , an increase of  $r_2$  could be achieved. This result could be due to many sharp edges on the surface of MNPs that leads to higher magnetic gradient.

**Table 2** Some studies information using cobalt ferrite as a MRI contrast agent in aqueous medium

| Reporter             | Sample name                                                                   | Size (nm) (TEM) | B, T | M <sub>s</sub> , emu/g | r <sub>1</sub> , s <sup>-1</sup> m M <sup>-1</sup> | r <sub>2</sub> , s <sup>-1</sup> m M <sup>-1</sup> | r <sub>2</sub> /r <sub>1</sub> |
|----------------------|-------------------------------------------------------------------------------|-----------------|------|------------------------|----------------------------------------------------|----------------------------------------------------|--------------------------------|
| Present study        | CoFe <sub>2</sub> O <sub>4</sub>                                              | 10.5            | 1.5  | 7.5                    | 1.15                                               | 58                                                 | 51                             |
| Vamvakidis-2018 [35] | CoFe <sub>2</sub> O <sub>4</sub> cluster                                      | 95 ± 12         | 1.5  | 116                    | 0.2                                                | 3.1                                                | 14.8                           |
| Iatridi-2016 [62]    | MnFe <sub>2</sub> O <sub>4</sub> :CoFe <sub>2</sub> O <sub>4</sub> -Copolymer | <100            | 1.5  |                        | 1.85                                               | 39.87                                              | 21.55                          |
| Sitthichai-2017      | CoFe <sub>2</sub> O <sub>4</sub>                                              | 20              | 1.5  | 40.36                  | —                                                  | 1313                                               | —                              |
|                      | CoFe <sub>2</sub> O <sub>4</sub> coated                                       | 12              | 1.5  | 99                     | —                                                  | 176                                                | —                              |
|                      | DMSA                                                                          |                 |      |                        |                                                    |                                                    |                                |
| Venkatesha-2014      | CoFe <sub>2</sub> O <sub>4</sub>                                              | 6,9,14          | 9.4  | 9.9, 34.2, 44.7        | —                                                  | 32, 2, 5                                           | —                              |
| Liu-2013             | CoFe <sub>2</sub> O <sub>4</sub> /H + /citrate                                | 12.98–37.89     | 0.47 | 28.42–45.50            | 12.8–27.8                                          | 16.6–75.1                                          | 1.3–2.7                        |
|                      | CoFe <sub>2</sub> O <sub>4</sub> /H + /citrate                                | 12.98–37.89     | 0.93 |                        | 7.1–9.1                                            | 12.4–51                                            | 1.75–5.6                       |
| Cheongwon- 2013      | CoFe <sub>2</sub> O <sub>4</sub>                                              | 10.5            | 4.7  | —                      | —                                                  | 66                                                 | —                              |
|                      | CoFe <sub>2</sub> O <sub>4</sub> @ SiO <sub>2</sub>                           | 15.8            |      | —                      | —                                                  | 179                                                | —                              |
|                      |                                                                               | 20              |      | —                      | —                                                  | 208                                                | —                              |
|                      |                                                                               | 24/2            |      | —                      | —                                                  | 302                                                | —                              |
|                      |                                                                               | 70.8            |      |                        |                                                    | 130                                                |                                |
| Ravichandran- 2015   | CoFe <sub>2</sub> O <sub>4</sub> nanowhisker                                  | 40–90           | 7    | 74                     | 2.367                                              | 256                                                | 108                            |
| Georgiadou-2015      | CoFe <sub>2</sub> O <sub>4</sub> -SBR                                         | 9–16            | 4    | 87                     | —                                                  | 167.4                                              | —                              |

In this study,  $r_1$  value was estimated at  $1.15 \text{ mM}^{-1} \text{ s}^{-1}$ . The longitudinal relaxivity,  $r_1$ , was related to the dead layer of paramagnetic material by the free spin on the surface of NPs.  $r_1$  also highly depended on the applied magnetic field. In this way, the increase of magnetic field strength decreases the value of  $r_1$  [49, 50]. Some reports indicated that for MNPs of below 20 nm,  $r_1$  increases with size [51].

The high value of  $r_2/r_1$  ratio indicated  $T_2$  contrast agent and vice versa [6, 52]. Some studies used cobalt ferrite as a contrast agent in the aqueous medium as shown in Table 2. According to the data of Table 2, the higher  $r_2/r_1$  was achieved in large sizes of MNPs and high magnetic field values. In this study, the calculated high  $r_2/r_1$  (51) compared to similar studies, indicated CoFe<sub>2</sub>O<sub>4</sub> MNPs as a good candidate for a negative contrast agent in clinical magnetic field strength. The optimal concentrations of MNPs for MR image were calculated for the clinical applications. The research result indicated that higher concentrations of 0.154 mM of MNPs did not have any clinical value to obtain higher signal decay. Recent studies have found that MFe<sub>2</sub>O<sub>4</sub> (M = Co, Ni, Cu or Zn) could induce the cytotoxicity and apoptosis by ROS generation and oxidative stress. The cytotoxicity of CoFe<sub>2</sub>O<sub>4</sub> nanoparticles and risks for biological systems must be checked because of their widespread application [53–56]. Few research studies reported the toxic potential of cobalt MNPs [57, 58]. However, there are studies indicating the lack of toxicity of cobalt ferrite nanoparticles [59–61]. The cytotoxicity study of CoFe<sub>2</sub>O<sub>4</sub> MNPs in this study indicated that concentrations of above 0.75 mM were toxic and the result agrees with the finding of Ravichandran *et al.* [14].

## 5 Conclusion

In this study, CoFe<sub>2</sub>O<sub>4</sub> MNPs were synthesised by a co-precipitation method. It successfully produced approximately small superparamagnetic cobalt ferrite MNPs by the average size of 10.45 nm.  $T_1$  and  $T_2$  relaxation times of hydrogen protons in aqueous solutions of varying concentrations were determined with a conventional MRI.  $T_1$  and  $T_2$  relaxivities ( $r_1$  and  $r_2$ ) were determined to be  $1.15$  and  $58 \text{ mM}^{-1} \text{ s}^{-1}$ , respectively. Owing to the high value of  $r_2/r_1$  (51), this research demonstrated the potential use of the synthesised MNPs as appropriate negative contrast agents at the conventional MRI system at low applied concentration. In addition, our results suggest that the CoFe<sub>2</sub>O<sub>4</sub> MNPs represent a perspective contrast agent suitable for cell labelling. The optimal concentration of CoFe<sub>2</sub>O<sub>4</sub> MNP as an MR contrast agent was obtained at 0.154 mM which is within a non-toxic concentration range.

In vitro cell viability assays indicate that the CoFe<sub>2</sub>O<sub>4</sub> MNPs showed no cellular viability reduction for concentrations up to 0.75 mM. As a result, it is suggested that this MNPs can be considered in further studies as a theranostic agent for improving the diagnostic and therapeutic application.

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